Use of tuberculin skin test, chest radiograph and interferon-gamma release assay to select migrants for treatment of latent tuberculosis

Michael G Flynn, Megan A Rees, Ayesha Saqib

# Abstract

Australian guidelines recommend the tuberculin skin test (TST) as the preferred test for latent tuberculosis infection (LTBI) with interferon-gamma release assay (IGRA) as a supplemental test to improve specificity. For many years the chest radiograph has been used to interpret a TST in the 5-9 mm range. The chest radiograph is known to predict subsequent reactivation of tuberculosis (TB). We report a retrospective database review of the Migrant Screening Clinic at Western Health Footscray Hospital during the period April 2010 to November 2011. Of 129 migrants under 35 years of age with TST 5-14 mm, IGRA was positive in 37 (28.7%). IGRA was positive in 7.7% of those with TST 5-9 mm and chest radiograph not suggestive of TB disease, 28.6% in those with TST 5-9 mm and chest radiograph that was suggestive of TB disease, and 39.5% in those with TST 10-14 mm (χ 2 trend=12.5, p=0.0004). There were 21 (16%) of 129 persons who had a negative IGRA but both TST >= 5 mm and a chest radiograph suggestive of TB disease. These data support existing recommendations to use the chest radiograph when interpreting a TST of 5-9 mm and recommending treatment of LTBI.

Keywords: screening; migrants; tuberculin; IGRA; latent tuberculosis

# Introduction

The detection and prevention of active TB developing after migration is a priority in both Victorian state and Australian national TB control strategies. 1,2 Australian guidelines recommend the tuberculin skin test (TST) as the preferred test for latent TB infection (LTBI) with interferon-gamma release assay (IGRA) as a supplemental test to improve specificity. 3 A recent Victorian study of TB contact tracing found the most common reason for performing IGRA was to increase the specificity of a TST in the 10-14 mm range.4

Australian research shows the annual incidence rate of TB in refugees increases with TST reaction size up to 30 mm, but is unrelated to bacille Calmette-Guérin (BCG) scar status.5 QuantiFERON-TB Gold is an interferon-gamma release assay (IGRA), which is known to be more specific for TB than the TST. 6 While some studies have suggested IGRA may have a better predictive value than tuberculin,7-9 others have not.10, 11 A recent review by the World Health Organization (WHO) found that in pooled estimates of the predictive utility of IGRA and TST, in head-to-head studies, the confidence intervals around risk ratio and incidence risk ratio for the TST and IGRA overlapped and were imprecise. 12 IGRA has been reported to have high negative predictive value in contact tracing studies,13-15 as well as screening studies in asylum seekers.16

Under arrangements with the Department of Health and Human Services Victoria, the Migrant Screening Clinic (MSC) in the Department of Respiratory and Sleep Disorders Medicine at Footscray Hospital Western Health has provided post-migration screening for TB in Victoria since 1996. Migrants who have been found to have an abnormal chest radiograph at the time of visa application are referred to the MSC by the Department of Immigration and Border Protection. 17 The MSC is a screening service, seeing each person once, and referring persons who need further assessment to other health services. 18, 19 Persons with symptoms suggestive of TB, or with one or more chest radiographs suggestive of active TB, are defined as suspected active TB and referred to other specialist clinics. The Victorian Government TB program is advised of non-attendees with a chest radiograph suggestive of active TB; TB program nurses are asked to contact the person, and may make a home visit. Those without such features, but with a history of TB diagnosis, a positive TST or IGRA, or one or more chest radiographs thought likely to represent previous TB, are defined as inactive TB.

Migrants are offered a TST if under 35 years of age and the chest radiograph is abnormal, or if under 35 years of age and a refugee,20 or if under 35 years of age and born in one of the high burden countries whose migrants have the highest TB incidence in Victoria.21, 22 The age of 35 years was chosen because the frequency and severity of isoniazid associated hepatitis increase with age.23 During the period under review (April 2010 to November 2011) chest radiographs were classified as described by Linh et al.24 Migrants with a TST of 15 mm or greater were offered referral for treatment for LTBI. If the TST was in the range 5-14 mm, a QuantiFERON-TB Gold assay (Cellestis, Australia) was performed. The result was reported as positive, negative or indeterminate according to the manufacturer’s guidelines. Those with a positive or indeterminate QuantiFERON-TB Gold were offered referral for consideration of treatment for LTBI.

We examine whether a comparison of TST and IGRA in our population supports the use of the chest radiograph to interpret TST.

# Methods

## Study population

In this observational study all persons under 35 years of age with TST and chest radiograph performed in the MSC during the period April 2010 to November 2011 were identified from a search of the Migrant Screening database. The information extracted included the chest radiograph classification, the results of TST and IGRA, and whether the migrant was referred for follow-up.

TST was considered positive according to the criteria of the American Thoracic Society (ATS),25 which are the criteria recommended in Victoria. 1 The ATS criteria are that a TST is positive at >= 10 mm, or at 5-9 mm if the chest radiograph demonstrates old fibrotic lesions thought to represent previous infection with TB. In this study, a TST of 5-9 mm was considered positive if the chest radiograph was classified as probable previous pulmonary TB disease (category 4a) or parenchymal infiltrate typical for TB (category 5a) as described by Linh et al.24

## Statistics

Proportion of those tested who were IGRA positive by TST category was examined using χ 2 for trend (Mantel extension, Epi Info v.6). Agreement between TST and IGRA was expressed as proportion in agreement, and corrected for chance agreement as Cohen’s kappa statistic κ with 95% confidence interval (CI).26

## Ethics approval

The study was approved by the Melbourne Health Human Research Ethics Committee and Western Health Office for Research.

# Results

In the 19 months between April 2010 and November 2011, a total of 2,430 persons attended the MSC for an initial assessment. There were 509 migrants under the age of 35 years with chest radiograph and TST. Eight persons with TST in the 5-14 mm range did not have IGRA performed and were excluded from analysis. Three persons with indeterminate IGRA were also excluded from analysis. Demographics, BCG status and TST results of the remaining 498 persons are shown in Table 1. There was no difference between mean TST in those with a BCG scar (15.4 mm) and those without a BCG scar (15.5 mm).

**Table 1. Demographics, BCG status and TST category**

| **Number of migrants 498** | |
| --- | --- |
| Females 45% | |
| Age (years) | |
| Mean  Median  Range | 26.5  26  16-34 |
| Country of Birth | |
| India | 36.5% |
| China | 15.3% |
| Nepal | 6.8% |
| Other | 38.8% |
| Unknown | 2.6% |
| Total | 100.0% |
| BCG scar  Present  Absent  Unknown | 46.8%  16.3%  36.9% |
| Total 100.0% | |
| TST category (n) | |
| 0-4 mm | 131 |
| 5-9 mm | 53 |
| 10-14 mm | 76 |
| >=15 mm | 238 |
| **Total** | **498** |

IGRA was performed in 129 migrants whose TST was in the range 5-14 mm. IGRA results after stratifying the TST 5-9 mm category by chest radiograph findings are shown in table 2. IGRA was positive in 7.7% of those with TST 5-9 mm and chest radiograph not suggestive of TB disease, 28.6% in those with TST 5-9 mm and chest radiograph that was suggestive of TB disease, and 39.5% in those with TST 10-14 mm (χ 2 trend=12.5, p=0.0004). When TST of 10-14 mm is also stratified by chest radiograph findings, there is relatively little difference in the proportion positive by IGRA: 38.6% in those with a chest radiograph not suggestive of TB disease, 42.1% in those with a chest radiograph that was suggestive of TB disease (table 3, χ 2 trend=11.8, p=0.0006).

**Table 2. Proportion IGRA positive by TST category, stratified by chest radiograph for TST 5-9 mm, as recommended by ATS.**

| **TST Category** | **n** | **IGRA negative n** | **IGRA positive n** | **IGRA**  **positive (row %)** |
| --- | --- | --- | --- | --- |
| 5-9 mm without radiographic disease | 39 | 36 | 3 | 7.7\* |
| 5-9 mm with radiographic disease | 14 | 10 | 4 | 28.6\* |
| 10-14 mm | 76 | 46 | 30 | 39.5\* |
| **Total** | **129** | **92** | **37** | **28.7** |

\* χ 2 trend=12.5, p=0.0004

**Table 3. Proportion IGRA positive by TST category, stratified by chest radiograph for both TST 5-9 and 10-14 mm categories.**

| **TST Category** | **n** | **IGRA negative n** | **IGRA positive n** | **IGRA**  **positive (row %)** |
| --- | --- | --- | --- | --- |
| 5-9 mm without radiographic disease | 39 | 36 | 3 | 7.7\* |
| 5-9 mm with radiographic disease | 14 | 10 | 4 | 28.6\* |
| 10-14 mm without radiographic disease | 57 | 35 | 22 | 38.6\* |
| 10-14 mm with radiographic disease | 19 | 11 | 8 | 42.1\* |
| **Total** | **129** | **92** | **37** | **28.7** |

\* χ2 trend=11.8, p=0.0006

When TST was considered positive at >= 10 mm, the proportion in agreement with IGRA was 59% (95%CI 50-67), Cohen’s kappa statistic κ = 0.24 (0.08-0.39). When TST was also considered positive at 5-9 mm if the chest radiograph showed probable previous pulmonary TB disease (category 4a) or parenchymal infiltrate typical for TB (category 5a), proportion in agreement was 54% (46-63), κ = 0.22 (0.07-0.36). In comparison to a 10 mm cut-off, use of the ATS criteria resulted in 14 persons changing from TST negative to positive, 10 of whom were IGRA negative and 4 of whom were IGRA positive (table 4).

**Table 4. Agreement between TST and IGRA for different criteria of positive TST**

|  | **TST positive if >=10 mm** | | **TST positive if >=10 mm,**  **or if 5-9 mm and radiographic disease** | |
| --- | --- | --- | --- | --- |
| **n** | **%** | **n** | **%** |
| TST negative - IGRA negative | 46 | 35.7 | 36 | 27.9 |
| TST negative - IGRA positive | 7 | 5.4 | 3 | 2.3 |
| TST positive - IGRA negative | 46 | 35.7 | 56 | 43.4 |
| TST positive - IGRA positive | 30 | 23.3 | 34 | 26.4 |
| **Total** | **129** | **100** | **129** | **100** |

Using the ATS criteria there were 328 persons with positive TST, comprising 14 with TST 5-9 mm plus fibrotic lesions on chest radiograph, 76 with TST 10-14 mm and 238 with TST >= 15 mm. If supplemental IGRA is used to interpret TST in the 5-14 mm range there were 275 persons with LTBI, comprising 37 with TST 5-14 mm plus positive IGRA, and 238 with TST >= 15 mm in whom IGRA was not performed. Of those 275 persons 264 (96%) were referred for follow-up. There were 21 (16%) of 129 persons who had a negative IGRA but both TST >= 5 mm and a chest radiograph considered consistent with fibrotic TB disease, of whom 6 (29%) were referred for follow-up.

# Discussion

In this study of migrants with a TST of 5-14 mm the presence of a chest radiograph suggestive of TB disease made a positive IGRA more likely, probably because the combination of TST and suggestive chest radiograph is more specific for LTBI than a positive TST with normal radiograph. Our data show that a chest radiographic finding of TB disease has a large effect on the chance of a positive IGRA when TST is 5-9 mm (28.6% versus 7.7%), but little effect when TST is 10-14 mm (42.1 versus 38.6%). Although proportion in agreement was not improved when TST was considered positive in the presence of radiographic TB disease, the χ 2 for trend analysis is highly significant and supports the ATS recommendation to use the chest radiograph when interpreting a TST of 5-9 mm.25 Given that a chest radiograph suggestive of previous TB is known to predict subsequent active TB,24 we believe that the chest radiograph should remain important to interpreting the TST, and in considering whether to recommend treatment of LTBI.

A direct measurement tool for LTBI in humans is currently unavailable.12 The presence of mycobacteria capable of causing reactivation TB cannot be directly detected by current methods such as bacterial culture, amplification of genomic material or detection of expressed proteins. Instead, evidence of their presence must be deduced from a specific immune response (TST, IGRA), or observing historical targeted inflammatory responses such as chest radiograph abnormalities, or by observation over time until reactivation disease occurs. This epidemiological information has been combined to determine estimates of TB reactivation in individuals. 27 Incorporating the chest radiograph in the assessment of likelihood of TB reactivation is particularly pertinent in patients who may have an attenuated immune response.28

Criteria for the diagnosis of LTBI in migrants from high incidence countries vary. In 1999, the ATS recommended targeted testing of persons at high risk for developing TB, and that a TST was positive at 10 mm or more in recent immigrants from high prevalence countries, or at 5 mm or more in persons with a chest radiograph demonstrating fibrotic lesions thought to represent previous TB disease.25 We do not use BCG status in the interpretation of TST because the ATS recommendation is that a positive TST in BCG-vaccinated persons indicates TB infection when the person tested is at increased risk for recent infection;25 and because a cohort study of Australian refugees found the incidence of active TB increased with the size of the TST, but that BCG status was not significant in multi-variate analysis.5 In 2006, the United Kingdom’s National Institute for Health and Clinical Excellence (NICE) recommended treatment for LTBI for people aged 35 or younger with a positive TST (6 mm or greater if no previous BCG, or 15 mm with previous BCG), and a positive interferon-gamma test (if this test was available).29 In 2010, the United States’ Centers for Disease Control and Prevention recommended that an IGRA may be used in place of (but not in addition to) a TST in all situations. 30 In 2011 the NICE guideline was revised to recommend initial IGRA, or initial TST with IGRA if TST 6 mm or greater. 31 In 2012, the National TB Advisory Committee Australia (NTAC) recommended TST and supplemental IGRA assessment for people with a positive TST.3 Although the NTAC guideline does not directly cite the ATS guidelines on reading the TST, it does cite Lee and Holzman,32 who do recommend the ATS criteria. In 2015 the WHO recommended that either TST or IGRA can be used to test for LTBI in high income and upper middle income countries with estimated TB incidence less than 100 per 100,000.12 The WHO guidelines note that chest radiography can be used in the algorithm to detect LTBI where screening is also directed towards active case finding. In 2016, the NICE guideline was revised again, and recommended initial TST, offer treatment if TST 5 mm or larger, and only offer IGRA if TST is unavailable.33 The 2016 NICE guidelines do not use the chest radiograph to interpret the TST. Also in 2016, the Australasian Society for Infectious Diseases (ASID) recommended using TST or IGRA to screen for LTBI; and that TST is positive at 10mm in those from refugee-like backgrounds, or 5 mm in the setting of severe malnutrition, HIV infection, immuno-suppression, or in children who are recent contacts of active TB cases. 20 While the ASID recommendations note the importance of chest radiograph in the detection of active TB, they do not mention incorporation of chest radiograph findings in the interpretation of TST. Whilst there are robust historical data to support the role of chest radiograph in predicting later reactivation of TB, it requires specialized skills to do so. 24

Many studies have compared TST with IGRA.30 Nienhaus et al.34 found agreement between QuantiFERON-TB Gold and TST was best with a TST diameter of at least 15 mm as cut-off point (89.8%); but the kappa value was best with at least 10 mm as cut-off point for the TST (0.37). Although TST, IGRA and chest radiograph have all been studied in the detection of latent TB, 35 we are unaware of any studies comparing TST and IGRA using the ATS recommendation that TST be considered positive at 5-9 mm if the chest radiograph suggests previous TB disease.25

The purpose of the Australian Government’s health requirement for immigrants is to protect the Australian community from public health and safety risks, particularly active TB.36 If active TB is found, Australian migration law does not allow a visa to be granted until the person has undergone treatment and been declared free of active TB.37 Applicants aged 11 years or older must undergo a chest radiograph,37 which is available when interpreting the TST.

# Conclusion

In migrants with a TST of 5-9 mm the presence of a chest radiograph suggestive of previous TB disease makes it more likely that IGRA will be positive. The chest radiograph is known to predict risk of subsequent active TB, and is recommended in the reading of the TST. Therefore the chest radiograph remains an important factor in considering whether to recommend treatment of LTBI.

# Acknowledgements

Michael Flynn contributed to conception and design, acquisition of data, analysis and interpretation of data, drafting the article and final approval. Megan Rees contributed to acquisition of data, revising the article and final approval. Ayesha Saqib contributed to conception and design, acquisition of data, and final approval.

# Author details

Dr Michael G Flynn, Physician, Department of Respiratory and Sleep Disorders Medicine, Footscray Hospital, Western Health, Footscray, Victoria. Dr Megan A Rees, Physician, Department of Respiratory and Sleep Disorders Medicine, Footscray Hospital, Western Health, Footscray, Victoria. Clinical Senior Lecturer, Melbourne Medical School, University of Melbourne, Parkville, Victoria. Dr Ayesha Saqib, Oncologist, The Epworth Centre, Level 5 Suite 6, 32 Erin Street, Richmond, Victoria. Corresponding author: Dr Michael Flynn, Department of Respiratory and Sleep Disorders Medicine, Footscray Hospital, Western Health, Footscray VIC 3011. Telephone: +61 3 8345 6169. Facsimile: +61 3 9318 6342. E-mail: michael.flynn@wh.org.au

# References

1. Aboltins C, Brown L, Curtis N, Darby J, Denholm J, Flynn M, et al. for the State of Victoria. Management, control and prevention of tuberculosis. Guidelines for health care providers. Melbourne, Australia: Victorian Government, 2015.
2. National Tuberculosis Advisory Committee of Communicable Diseases Network Australia. The strategic plan for control of tuberculosis in Australia: 2011–2015. Commun Dis Intell 2012;36(3):E286-293.
3. National Tuberculosis Advisory Committee. Position statement on interferon-γ release assays in the detection of latent tuberculosis infection. Commun Dis Intell 2012;36(1):125-131.
4. Goebel KM, Tay EL, Denholm JT. Supplemental use of an interferon-gamma release assay in a state-wide tuberculosis contact tracing program in Victoria: a six year review. Commun Dis Intell 2015;39(2):E191-196.
5. Marks GB, Bai J, Simpson SE, Sullivan EA, Stewart GJ. Incidence of Tuberculosis among a Cohort of Tuberculin-Positive Refugees in Australia - Reappraising the Estimates of Risk. Am J Respir Crit Care Med 2000; 162:1851–1854.
6. Lalvani A, Pareek M. Interferon gamma release assays: principles and practice. Enferm Infecc Microbiol Clin 2010; 28:245-252.
7. Diel R, Loddenkemper R, Meywald-Walter K, Niemann S, Nienhaus A. Predictive value of a whole blood IFN-g assay for the development of active tuberculosis disease after recent Infection with Mycobacterium tuberculosis. Am J Respir Crit Care Med 2008; 177:1164–1170.
8. Doherty TM, Demissie A, Olobo J, Wolday D, Britton S. Eguale T, Ravn P, Andersen P. Immune responses to the Mycobacterium tuberculosis-specific antigen ESAT-6 signal subclinical infection among contacts of tuberculosis patients. J Clin Microbiol 2002; 40:704–706.
9. Higuchi K, Harada N, Mori T, Sekiya Y. Use of QuantiFERON®-TB Gold to investigate tuberculosis contacts in a high school. Respirology 2007; 12:88–92.
10. Bakir M, Millington KA, Soysal A, Deeks JJ, Efee S, Aslan Y, Dosanjh DPS, Lalvani A. Prognostic value of a T-cell–based, interferon-gamma biomarker in children with tuberculosis contact. Ann Intern Med 2008;149:777-786.
11. Hill PC, Jackson-Sillah DJ, Fox A, Brookes RH, de Jong BC, Lugos MD, et al. Incidence of tuberculosis and the predictive value of ELISPOT and Mantoux tests in Gambian case contacts. PLoS ONE 3(1): e1379. doi:10.1371/journal.pone.0001379
12. World Health Organization. Guidelines on the management of latent tuberculosis infection. Geneva, Switzerland: World Health Organization, 2015.
13. Aichelburg MC, Rieger A, Breitenecker F, Pfistershammer K, Tittes J, Eltz S, Aichelburg AC, Stingl G, Makristathis A, Kohrgruber N. Detection and prediction of active tuberculosis disease by a whole blood interferon-gamma release assay in HIV1-infected individuals. Clin Infect Dis 2009; 48:954–962
14. Altet N, Dominguez J, de Souza-Galvão M-L, M. Angeles Jimenez-Fuentes M, Mila C, Solsona J, Soriano-Arandes A, Latorre I, Lara E, Cantos A, Ferrer M, Orcau A, Ruiz-Manzano J, Cayla J. Predicting the Development of Tuberculosis with the Tuberculin Skin Test and QuantiFERON Testing. Ann Am Thorac Soc 2015;12(5):680-688.
15. Diel R, Loddenkemper R, Niemann S, Meywald-Walter K, Nienhaus A. Negative and Positive Predictive Value of a Whole-Blood Interferon-gamma Release Assay for Developing Active Tuberculosis. An update. Am J Respir Crit Care Med 2011(1);183: 88–95.
16. Harstad I, Winje B, Heldal E, Oftung F, Jacobsen G. Predictive values of QuantiFERON®-TB Gold testing in screening for tuberculosis disease in asylum seekers. Int J Tuberc Lung Dis 2010;14(9):1209–1211.
17. Australian Government. Department of Immigration and Border Protection . Fact Sheet - Health requirement. Available from https://www.border.gov.au/about/corporate/information/fact-sheets/22health, accessed 23 October 2016.
18. Flynn M, Brown L, Tesfai A, Lauer T. Post-migration screening for active tuberculosis in Victoria, Australia. Int J Tuberc Lung Dis 2012;16(1):50-54
19. Flynn MG, Brown LK. Treatment of latent tuberculosis in migrants to Victoria. Commun Dis Intell 2015;39(4):E578–E583
20. Australasian Society for Infectious Diseases and Refugee Health Network of Australia. Guidelines writing group. Recommendations for comprehensive post-arrival health assessment for people from refugee-like backgrounds, 2nd ed, pp.25-30. Australasian Society for Infectious Diseases, Surry Hills, NSW, 2016.
21. MacIntyre CR, Dwyer B, Streeton JA. The epidemiology of tuberculosis in Victoria. Med J Aust 1993;159:672– 677.
22. Dye C, Scheele S, Dolin P, Pathania V, Raviglione MC. Global burden of tuberculosis: estimated incidence, prevalence and mortality by country. JAMA 1999;282:677–686.
23. Saukkonen JJ, Cohn DL, Jasmer RM, Schenker S, Jereb JA, Nolan CM, et al. An official ATS statement: Hepatotoxicity of antituberculosis therapy. Am J Respir Crit Care Med 2006;174(8):935–952.
24. Linh NN, Marks GB, Crawford ABH. Radiographic predictors of subsequent reactivation of tuberculosis. Int J Tuberc Lung Dis 2007; 11:1136–1142.
25. American Thoracic Society. Targeted tuberculin testing and treatment of latent tuberculosis infection. Am J Respir Crit Care Med 2000; 161:S221–S247.
26. Cohen JL. A coefficient of agreement for nominal scales. Educ Psch Meas 1960; 20 (1):37-46.
27. Menzies M, Gardiner G, Farhat M, Greenaway C, Pai M. Thinking in three dimensions: a web-based algorithm to aid the interpretation of tuberculin skin test result. Int J Tuberc Lung Dis 2008;12(5):498–505.
28. Piccazzo R, Paparo F, Garlaschi G. Diagnostic accuracy of chest radiography for the diagnosis of tuberculosis (TB) and its role in the detection of latent TB infection: a systematic review. J Rheumatol 2014;91;32-40.
29. National Institute for Health and Clinical Excellence. Clinical Guideline 33. Clinical diagnosis and management of tuberculosis, and measures for its prevention and control. London, United Kingdom: 2006
30. Centers for Disease Control and Prevention. Updated guidelines for using interferon gamma release assays to detect Mycobacterium tuberculosis infection. Morb Mortal Wkly Rep 2010;59(RR–5):1–25.
31. National Institute for Health and Clinical Excellence. Clinical Guideline 117. Tuberculosis. Clinical diagnosis and management of tuberculosis, and measures for its prevention and control. London: 2011.
32. Lee E, Holzman RS. Evolution and current use of the tuberculin test. Clinical Infectious Diseases 2002;34(3):365–70.
33. National Institute for Health and Care Excellence. Tuberculosis. NICE, 2016. Available from https://www.nice.org.uk/guidance/ng33/, accessed 23 October 2016.
34. Nienhaus A, Schablon A, Diel R. Interferon-gamma release assay for the diagnosis of latent TB infection - analysis of discordant results when compared to the tuberculin skin test. PLoS ONE 3(7): e2665. doi:10.1371/journal.pone.0002665
35. Jeong YJ, Yoon S, Koo HK, Lim HJ, Lee JS, Lee SM, Yang SC, Yoo CG, Kim YM, Han SK, Yim JJ. Positive tuberculin skin test or interferon-gamma release assay in patients with radiographic lesions suggesting old healed tuberculosis. J Korean Med Sci 2012;27(7):761-766.
36. Australian Government. Department of Immigration and Border Protection. Overview of the health requirement. Accessed 29 January 2017. Available from: https://www.border.gov.au/Trav/Visa/Heal/overview-of-the-health-requirement
37. Australian Government. Department of Immigration and Border Protection. Fact sheet - health requirement. Accessed 29 January 2017. Available from: https://www.border.gov.au/about/corporate/information/fact-sheets/22health



© Commonwealth of Australia 2018 - ISSN: 2209-6051 (Online)

This work is copyright. You may download, display, print and reproduce the whole or part of this work in unaltered form for your own personal use or, if you are part of an organisation, for internal use within your organisation, but only if you or your organisation do not use the reproduction for any commercial purpose and retain this copyright notice and all disclaimer notices as part of that reproduction. Apart from rights to use as permit­ted by the Copyright Act 1968 or allowed by this copyright notice, all other rights are reserved and you are not allowed to reproduce the whole or any part of this work in any way (electronic or otherwise) without first being given the specific written permission from the Commonwealth to do so. Requests and inquiries concerning reproduction and rights are to be sent to the Online, Services and External Relations Branch, Department of Health, GPO Box 9848, Canberra ACT 2601, or by email to copyright@health.gov.au

Communicable Diseases Intelligence aims to disseminate information on the epidemiology and control of communicable diseases in Australia. Communicable Diseases Intelligence invites contri­butions dealing with any aspect of communicable disease epidemiology, surveillance or prevention and control in Australia. Submissions can be in the form of original articles, short reports, surveil­lance summaries, reviews or correspondence. Instructions for authors can be found in Commun Dis Intell 2016;40(1):E189–E193.

Communicable Diseases Intelligence contributes to the work of the Communicable Diseases Network Australia.

http://www.health.gov.au/cdna

This journal is indexed by Index Medicus and Medline.

**Disclaimer:** Opinions expressed in Communicable Diseases Intelligence are those of the authors and not necessarily those of the Australian Government Department of Health or the Communicable Diseases Network Australia. Data may be subject to revision.

**Editor:** Cindy Toms  
**Deputy Editor:** Phil Wright  
**Editorial and Production Staff:** Leroy Trapani, Kasra Yousefi  
**Editorial Advisory Board:** Peter McIntyre (Chair), David Durrheim, Mark Ferson, John Kaldor, Martyn Kirk  
**Website:** http://www.health.gov.au/cdi

Communicable Diseases Intelligence is produced by Health Protection Policy Branch, Office of Health Protection, Australian Government, Department of Health, GPO Box 9848, (MDP 6) CANBERRA ACT 2601;

**Email:** cdi.editor@health.gov.au