Nocardiosis in the Illawarra-Shoalhaven Regions from 2010 to 2019

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Abstract

*Nocardia* species are a diverse group of gram-positive bacteria that are encountered as pathogens in clinical practice. They are ubiquitous environmental organisms classically causing opportunistic infections in immunocompromised individuals. This case series describes the epidemiological, microbiological and clinical features of 26 culture-proven cases of nocardiosis presenting to the public healthcare service in a regional area of New South Wales over a ten year period. These cases involved ten distinct species of *Nocardia* and appeared to demonstrate increasing incidence over time.

The majority of *Nocardia* isolates (88%) were grown on conventional bacteriological media and speciation was primarily via 16S rRNA sequencing and/or matrix assisted laser desorption ionisation—time of flight mass spectrometry. Similar to other Australian *Nocardia* studies, the isolates seen in this case series were universally susceptible to trimethoprim-sulfamethoxazole. This was reflected in treatment data, with all but one patient receiving this agent as part of their antibiotic regimen.

While similar case series have been reported from other Australian states, there are no published clinical studies of nocardiosis in New South Wales. Given the nature of *Nocardia* species as an environmental saprophyte, this study examined the association between incidence of nocardiosis and regional geographic or climatic variation but was unable to demonstrate a statistically significant correlation.

Keywords: Nocardia; nocardial infection; Australia; microbiology

Introduction

*Nocardia* species are a diverse group of filamentous gram-positive aerobic bacteria of the order Actinomycetales. They are most commonly associated with opportunistic infections, often disseminated, in immunocompromised patients. They can also cause localised infection following direct inoculation, usually without dissemination, in immunocompetent individuals.

These bacteria are ubiquitous within the environment and are present in soil, water and organic material. They have a variable geographic distribution. Data on infections in Australia is limited and comprised of case series and small observational studies. While previous reports have defined the pattern of nocardiosis in the Northern Territory, Queensland, Victoria, and the Australian Capital Territory, none have examined the presentations related to *Nocardia* infections in New South Wales (NSW).

The Illawarra-Shoalhaven region extends from Helensburgh on the southern outskirts of Sydney to North Durras on the NSW South Coast. This region had a population of approximately 400,000 in 2015; it comprises a diverse range of human environments from major metropolitan to rural areas. The Illawarra Shoalhaven Local Health District (ISLHD) provides public healthcare services to this region.
Wollongong Hospital is the tertiary referral hospital and location of the NSW Health Pathology microbiology laboratory serving ISLHD. In this study we sought to examine all cases of nocardiosis presenting to ISLHD services over the ten-year period from 2010 to 2019, to understand the local epidemiology, microbiology and clinical characteristics of *Nocardia* infections compared to that described in other studies. We also assessed the association between incidence of nocardiosis in ISLHD and regional rainfall patterns.

**Methods**

Cases of culture-confirmed nocardiosis were identified through interrogation of the NSW Health Pathology Wollongong Hospital microbiology laboratory records for the period 2010–2019. The NSW Health Pathology reference laboratories, Institute for Clinical Pathology and Medical Research (Westmead Hospital) and Australian National Reference Laboratory in Medical Mycology (Royal North Shore Hospital), were also canvassed for isolates sent to these sites from ISLHD during this time period. Additional cases were identified through a search of the clinical records of the Department of Infectious Diseases at Wollongong Hospital and Shoalhaven District Memorial Hospital.

Routine cultures of swabs and respiratory samples were inoculated, as a minimum, onto horse blood agar and MacConkey agar and incubated for two days. Tissue samples, normally sterile body fluids and positive blood culture subcultures were additionally inoculated onto chocolate agar and incubated routinely for three days. Direct samples were performed/ examined and blood agar plate cultures were incubated aerobically for seven days. Blood cultures were inoculated into BD BACTEC Plus Aerobic/P (Becton Dickinson) and processed with the BD BACTEC 9240 (Becton Dickinson) system with a routine incubation period of five days. If mycobacterial culture was requested the samples were processed by the NSW Health Pathology Mycobacteriology laboratory at St George Hospital, Kogarah, NSW Australia. Samples were inoculated onto two Lowenstein-Jensen agar slopes (with and without pyruvate) and a Mycobacteria Growth Indicator Tube (Becton Dickinson) and incubated routinely for eight weeks. No changes in the laboratory methods for *Nocardia* species culture were made during the study period.

Culture isolates were presumptively identified as *Nocardia* species based on strict aerobic growth, colony morphology, Gram stain morphology and positive modified acid-fast (Ziehl-Neelsen) stain and negative standard acid-fast (Ziehl-Neelsen) stain. All presumptive *Nocardia* isolates were forwarded to either of the two above-mentioned NSW Health Pathology reference laboratories for confirmation, species identification, and antimicrobial susceptibility testing. Species identification was made using 16S rRNA sequencing and/or matrix-assisted laser desorption ionisation–time of flight mass spectrometry (MALDI-TOF MS). Antimicrobial susceptibilities were determined using a commercially-available broth microdilution method, Sensititre™ RAPMYCO (Trek Diagnostic Systems, Biocentric), with the exception of meropenem susceptibility which was determined using Epsilometer test (E-Test®, bioMérieux). Where possible, susceptibilities were interpreted using Clinical and Laboratory Standards Institute (CLSI) breakpoints.

For all patients with cultures positive for *Nocardia* species, a search of the ISLHD electronic medical record was conducted to compile relevant data including patient demographics, risk factors for nocardiosis, clinical features of infection, treatment and outcome data.

Limited disease was defined as: either (i) skin and soft tissue infection with or without contiguous osteomyelitis, or (ii) pulmonary disease, both without evidence of other organ involvement. Disseminated disease was defined as disease involving two or more organ systems,
bacteraemia, or isolated central nervous system (CNS) disease. Survival calculations were based on documented evidence that a patient was alive or deceased. If no evidence was available regarding a patient after a given time point, they were excluded from analysis at that time point. Local incidence was calculated using the number of nocardiosis cases over the ten-year study period and postcode population data from the 2016 Australian Census, expressed as cases per 100,000 persons. Case numbers were compared to Bureau of Meteorology total annual rainfall data for Jamberoo (a geographically-central area of the region). Statistical correlation between rainfall and case numbers was compared using a Pearson product-moment linear regression coefficient. Statistical testing was then performed based on transforming this coefficient into a Student’s T statistic, based on a presumption that both variables were from an uncorrelated bivariate normal distribution, which was itself tested using a Shapiro-Wilk test for each variable.

The data were deidentified and aggregated with analysis of descriptive statistics performed using the Microsoft Excel platform. The study underwent ethics approval through the joint University of Wollongong/Illawarra Shoalhaven Local Health District Human Ethics Review Committee (approval number: ETH10680).

Results

Epidemiology and clinical features

Twenty-six cases of culture-proven Nocardia sp. infection were identified. Patient demographic data and predisposing conditions are displayed in Table 1. Since 2010, cases of nocardiosis presenting to ISLHD facilities involved ten distinct species (Table 2), resulting in a variety of clinical presentations including skin and soft tissue infection with or without osteomyelitis (N = 8), pulmonary disease (N = 8) or disseminated disease (N = 9) (Figure 1). One case had limited clinical data available and was excluded from the treatment and outcomes analyses. Two cases had incomplete antimicrobial susceptibility data (with one of these having no available antimicrobial susceptibilities). No patients included in the study were receiving antibiotic prophylaxis with activity against Nocardia species (e.g. trimethoprim-sulfamethoxazole) prior to presentation. Median and range displayed for prednisolone dose was calculated from data available in the medical record; doses were unavailable for three patients.

While pulmonary and disseminated disease were caused by several different Nocardia species (Figure 2), locally-acquired skin and soft tissue infection was caused almost exclusively by N. brasiliensis. One patient experienced skin and soft tissue infection with contiguous osteomyelitis due to N. rhamnosiphila acquired by traumatic inoculation sustained overseas; another case of soft tissue infection caused by N. thailandica was again related to direct inoculation.

Pulmonary imaging findings

Twelve patients had pulmonary involvement (either isolated or in the setting of disseminated disease). Chest computerised tomography (CT) reports were available for analysis of the imaging findings in 11 patients. Chest CT in 2/11 (18%) patients demonstrated single nodules and in 5/11 (45%) demonstrated multiple nodules; in 3/7 patients with pulmonary nodules there was cavitation. Consolidation was present on imaging in 3/11 (27%) patients, of whom two had an associated pleural effusion. The remaining patient had mediastinal lymphadenopathy without lung nodules or consolidation.

Local epidemiology

Local incidence data to postcode level (Figure 3) demonstrates that the highest rates of nocardiosis (approximately 19 cases per 100,000 persons over the 10 year study period) occurred in the rural southern Shoalhaven region as well as in the postcodes around Lake Illawarra (just south of Wollongong).
Table 1: Demographic information and epidemiological risk factors

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Number of patients</th>
<th>%</th>
</tr>
</thead>
<tbody>
<tr>
<td>Female</td>
<td>10/26</td>
<td>39</td>
</tr>
<tr>
<td>Median age in years [Range]</td>
<td>67 [11–92]</td>
<td></td>
</tr>
<tr>
<td><strong>Risk factors</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Haematological malignancy</td>
<td>5/26</td>
<td>19</td>
</tr>
<tr>
<td>Corticosteroid therapy:</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Median dose (mg/dy prednisolone) [Range]</td>
<td>11/26</td>
<td>12.5</td>
</tr>
<tr>
<td></td>
<td></td>
<td>[7.5–25]</td>
</tr>
<tr>
<td>Other immunosuppression</td>
<td>6/26</td>
<td>23</td>
</tr>
<tr>
<td>Chronic lung disease</td>
<td>5/26</td>
<td>19</td>
</tr>
<tr>
<td>Type 2 diabetes mellitus</td>
<td>5/26</td>
<td>19</td>
</tr>
<tr>
<td>End stage renal disease on haemodialysis</td>
<td>1/26</td>
<td>4</td>
</tr>
<tr>
<td>Renal transplant</td>
<td>3/26</td>
<td>12</td>
</tr>
<tr>
<td>HIV</td>
<td>1/26</td>
<td>4</td>
</tr>
<tr>
<td>Trauma/inoculation</td>
<td>3/26</td>
<td>11</td>
</tr>
<tr>
<td>No identified risk factor</td>
<td>3/26</td>
<td>11</td>
</tr>
</tbody>
</table>

Figure 1: Incident cases of culture-proven nocardiosis presenting to the Illawarra Shoalhaven Local Health District
Table 2: Incident cases of culture proven nocardiosis presenting to the Illawarra Shoalhaven Local Health District classified by Nocardia species during the ten-year study period

<table>
<thead>
<tr>
<th>Species</th>
<th>Number of isolates</th>
</tr>
</thead>
<tbody>
<tr>
<td>Nocardia brasiliensis</td>
<td>6</td>
</tr>
<tr>
<td>Nocardia farcinica</td>
<td>6</td>
</tr>
<tr>
<td>Nocardia cyriacigeorgica</td>
<td>5</td>
</tr>
<tr>
<td>Nocardia nova</td>
<td>3</td>
</tr>
<tr>
<td>Nocardia paucivorans</td>
<td>1</td>
</tr>
<tr>
<td>Nocardia vinacea</td>
<td>1</td>
</tr>
<tr>
<td>Nocardia sp. (N. gamkensis or N. exalbida)</td>
<td>1</td>
</tr>
<tr>
<td>Nocardia beijingensis</td>
<td>1</td>
</tr>
<tr>
<td>Nocardia rhamnosiphila</td>
<td>1</td>
</tr>
<tr>
<td>Nocardia thailandica</td>
<td>1</td>
</tr>
</tbody>
</table>

Annual nocardiosis case numbers in the Illawarra-Shoalhaven region were compared to average annual rainfall in Jamberoo (Figure 4) to examine for any possible linkage between incidence of infection and climatic factors. This had a correlation coefficient of – 0.34 (p = 0.2) and thus did not demonstrate statistically significant linkage.

**Microbiology**

Twenty-five separate microbiological samples were processed through the NSW Health Pathology Wollongong Hospital laboratory for the 26 cases. Two cases had isolates through external pathology providers that were not available for analysis. Twelve of the 25 samples (48%) were from sterile sites (blood, tissue, pleural fluid or deep operative aspirate) while 4/25 samples (16%) were sputum and 9/25 (36%) were superficial swabs or pus. Direct microscopy with Gram stain, modified acid-fast and standard acid-fast stains was performed on...
23/25 samples on either the day of sample collection or the following day. In 10/23 samples (43%), organisms consistent with *Nocardia* species were seen on this examination.

Twenty-two of the 25 isolates (88%) grew on conventional bacteriological media as described in the methods above. Eleven of the 25 culture-positive samples (44%) were inoculated onto mycobacterial culture media (Lowenstein Jensen media) of which 3/11 (37%) grew the *Nocardia* isolate. In all three of these samples *Nocardia* grew solely on mycobacterial culture media. The median time from collection to growth detection for all isolates was three days; however, this was skewed by isolates grown
only on mycobacterial culture media, which grew after a median of 34 days as opposed to those isolates cultured on routine bacteriological media which had a median time to growth detection of two days. *Nocardia* species were isolated from blood cultures in two cases.

Species identification was determined using 16S rRNA sequencing in 10/25 samples (40%), MALDI-TOF MS in 10/25 (40%) and using both methods in 5/25 (20%).

**Antibiograms**

Antimicrobial susceptibility testing of isolates showed all were susceptible to trimethoprim-sulfamethoxazole, amikacin and, where examined (n = 11), linezolid; however, a high proportion were resistant to ciprofloxacin and carbapenems (Figure 5). Analysis of resistance patterns for species with more than one isolate (Table 3) demonstrates significant variability in resistance patterns between isolates (especially to ciprofloxacin and amoxicillin-clavulanate) even within the former *N. asteroides* complex.

**Antimicrobial therapy**

Data relating to treatment were available for 25 of the 26 cases identified. Initial therapy was with a single antimicrobial agent in 28% (7/25), two antimicrobials in 52% (13/25), and three antimicrobials in 20% (5/25) of cases. Twenty-four of 25 patients (96%) were treated with trimethoprim-sulfamethoxazole following identification of *Nocardia* sp. in diagnostic samples. The most commonly used agents, in addition to trimethoprim-sulfamethoxazole as part of an initial empiric combination therapy, were a carbapenem (61%; 11/18), ceftriaxone (27%; 5/18) and ciprofloxacin (16%; 3/18). This initial combination therapy was given for a median duration of ten days in skin and soft tissue disease, 28 days in pulmonary disease and...
Figure 5: In vitro antibiogram for all *Nocardia* isolates included in the study with antimicrobial susceptibility available (N = 25)*a*

![Antibiogram](image)

*a All isolates had antibiotic susceptibility testing for trimethoprim-sulfamethoxazole, ciprofloxacin, amikacin, imipenem, amoxicillin-clavulanate, minocycline and ceftriaxone. Only 7 isolates had susceptibility testing for meropenem reported and 11 isolates had susceptibility testing for linezolid performed.

six weeks in the case of disseminated or central nervous system disease. Monotherapy was most commonly trimethoprim-sulfamethoxazole (85%; 6/7).

Consolidation therapy (defined as a long-term maintenance or eradication treatment to prevent relapse) was with a single drug in 64% of cases (16/25) and with dual therapy in 36% (9/25). The single agent used most commonly was trimethoprim-sulfamethoxazole (88%; 14/16). All dual-therapy regimens contained trimethoprim-sulfamethoxazole with, most commonly, the addition of either a tetracycline (doxycycline or minocycline) (56%; 5/9) or moxifloxacin (33%; 3/9). This consolidation therapy was given for a median duration of three months in skin and soft tissue disease, six months in pulmonary disease, and 12 months in disseminated or CNS disease.

Overall, 44% (11/25) of the patients with data available had documented evidence of an adverse drug reaction. This comprised acute kidney injury in 16% of cases (4/25); gastrointestinal disturbance in 12% (3/25); liver function derangement in 8% (2/25); and cytopenia in 8% (2/25).

**Mortality statistics**

One-month mortality was 3.8% (1/26), whereas six-month mortality and 12-month mortality were 9.1% (2/22) and 15.7% (3/19) respectively. Only one patient died during initial therapy. No deaths were observed in the cohort with skin and soft tissue disease; however, several of these patients were lost to follow up.

**Discussion**

This retrospective study reports the demographic, clinical and microbiological
Table 3: Antibiotic susceptibility by species, for species with more than one isolate included in the study

| Isolate species | (number of isolates) | Ceftriaxone |  |  |  | Minocyclin |  |  |  | Amoxicillin-clavulanate |  |  |  | Imipenem |  |  |  | Amikacin |  |  |  | Ciprofloxacin |  |  |  | Trimethoprim-sulfamethoxazole |  |  |  |  |
|-----------------|----------------------|-------------|---|---|---|-------------|---|---|---|------------------------|---|---|---|-------------|---|---|---|-----------|---|---|---|----------------|---|---|---|-------------------|---|---|---|------------------|---|---|---|-------------------|
|                 |                      | Resistant % | Intermediate % | Susceptible % |  | Resistant % | Intermediate % | Susceptible % |  | Resistant % | Intermediate % | Susceptible % |  | Resistant % | Intermediate % | Susceptible % |  | Resistant % | Intermediate % | Susceptible % |  | Resistant % | Intermediate % | Susceptible % |  | Resistant % | Intermediate % | Susceptible % |  |  |
| N. brasiliensis  | (6)                  | 17          | 17             | 67            | 0 | 50           | 50            |             | 0 | 17          | 83            |             | 100          | 0             | 0             | 100          | 83          | 17            | 0             | 0             | 100          | 0             | 100          | 0             | 100          | 0             | 100          | 0             | 100          | 0             | 0           | 100          | 0             | 0             | 100          | 0             | 0             | 100          |
| N. cyriacigeorgica | (4)                | 25          | 0              | 75            | 0 | 75           | 25            | 25          | 25 | 75          | 0              | 0             | 25           | 0              | 100          | 75           | 0             | 25          | 0             | 25           | 75           | 0              | 100          | 0             | 100          | 0             | 0             | 100          | 0             | 0             | 100          |
| N. farcinica     | (6)                  | 100         | 0              | 0             | 0 | 80           | 20            | 0           | 20 | 80          | 17            | 17            | 0             | 100          | 0             | 100          | 0             | 40           | 60            | 0             | 100          | 0             | 100          | 0             | 0             | 100          | 0             | 0             | 100          |
| N. nova          | (3)                  | 0           | 33             | 67            | 0 | 100          | 0             | 100         | 0   | 100         | 0              | 0             | 100          | 0             | 100          | 0             | 100          | 0             | 100          | 0             | 100          | 0             | 100          | 0             | 100          | 0             | 100          | 0             | 100          |

a Due to the low numbers of susceptibility testing to meropenem and linezolid within each species, these antibiotics are excluded from this analysis.
characteristics of 26 patients with culture-proven *Nocardia* infection over a ten-year period in a large and geographically diverse health service in regional NSW.

Comparison with other large Australian cohort studies demonstrates regional variation in the demographic characteristics and risk profile of patients with nocardiosis. Our cohort had similar demographic features to other studies of patients with nocardiosis in southern and eastern Australia, whereas nocardiosis patients in the Northern Territory were more likely to be younger, female, and of Aboriginal or Torres Strait Islander heritage.

The majority of patients in this study (58%) were immunocompromised, most commonly due to corticosteroid use (42%), non-corticosteroid immunosuppressive medication (23%) and/or haematological malignancy (19%); solid organ transplantation was identified as a risk factor in just three patients in our cohort (12%). By comparison, fewer than half of the patients in similar Australian nocardiosis studies were considered immunocompromised. High rates of chronic disease, including chronic lung disease, diabetes mellitus and end-stage renal disease, are commonly reported in nocardiosis studies; however, our study is notable for a much lower proportion of patients with chronic lung disease than other Australian nocardiosis studies (19% vs > 50%).

Given the variation in demographics and risk profile, it is unsurprising that the clinical characteristics of infections due to *Nocardia* sp. were also quite different. Paige and Spelman reported on 67 patients with nocardiosis, of whom 53 (79%) had isolated pulmonary disease and just eight (12%) had disseminated disease. In contrast, eight patients in our study (31%) had isolated pulmonary disease and nine (35%) had disseminated disease. This likely reflects the above-mentioned lower proportion of patients with chronic lung disease in our study and the higher proportion of patients with immunocompromise. Furthermore, in approximately one-third of patients in our study, nocardiosis manifested as isolated skin and soft tissue infection, often with a history of trauma at the site that may have resulted in inoculation of environmental organisms; all of which had a clinical syndrome consistent with *Nocardia* sp. infection. In contrast, only 7% of patients in the Victorian study had *Nocardia* infection limited to soft tissues and bone.

Antimicrobial susceptibility testing results in our study showed universal susceptibility of *Nocardia* sp. to trimethoprim-sulfamethoxazole and, where examined, linezolid, in keeping with other laboratory studies from NSW. This finding provides additional support for current Australian guidelines for the antimicrobial treatment of *Nocardia* sp. infection. It is also consistent with our treatment data showing that trimethoprim-sulfamethoxazole formed the backbone of most therapeutic regimens.

It should be noted that as ISLHD serves a large geographical area with a wide range of terrain and population density and that the majority of cases occurred in patients living away from the more densely populated centres in the northern Illawarra.

The majority of cases of nocardiosis reported here (18/26) were identified in the last four years of the ten-year study, coinciding with a significant drought event in south-eastern Australia. Other Australian studies have hypothesised that the incidence of nocardiosis may be influenced by climatic conditions. In addition, international evidence suggests an association with dry, warm climates, for example in the south-eastern United States of America. Our study was unable to demonstrate a statistically significant correlation between the incidence of nocardiosis in ISLHD and regional rainfall trends; however, the study was likely underpowered for this purpose. In addition, for a large geographic area, complete rainfall data was only available for a single, albeit central, area of the region. This limited the resolution of analysis.
This study was limited in its ability to sufficiently resolve trends in nocardiosis epidemiology by the small sample size. Furthermore, the limitations of retrospective data collection from clinical records (particularly with regards to treatment data) prevented more extensive analysis of the rationale and timing of changes to the treatment regimens for this uncommon disease.

We hypothesise that the increasing prevalence of patients with chronic comorbidities and immunocompromise, living in rural and regional communities exposed to environmental sources of *Nocardia* sp., may lead to an increased incidence of nocardiosis in the Illawarra-Shoalhaven region and that comparable trends may be seen in other regions with similar demographics and epidemiology. Larger prospective studies are required to test this hypothesis. The association between climatic conditions and incidence of nocardiosis remains unproven, but given the potential implications of global climate change, this concept warrants further investigation.

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